



Course Syllabus: Molecular & Cellular Biology - B 241

Division	Biological and Environmental Sciences & Engineering Division
Course Number	B 241
Course Title	Molecular & Cellular Biology
Academic Semester	Fall
Academic Year	2018/2019
Semester Start Date	08/26/2018
Semester End Date	12/11/2018
Class Schedule (Days & Time)	02:30 PM - 06:00 PM Mon Thu

Instructor(s)				
Name	Email	Phone	Office Location	Office Hours
Samir Hamdan	samir.hamdan@kaust.edu.sa	+966128082384		Tuesday: 10:00-11:00 am Building 2 Room 4221
Samah Zeinab Gadhoum	Samah.Gadhoum@kaust.edu.sa	+966128082930		

Teaching Assistant(s)	
Name	Email
Dr. Samah Zeineb Gadhoum; Building 9, third floor and room 3354	samah.gadhoum@kaust.edu.sa

Course Information	
Comprehensive Course Description	The course provides first hands-on experience in a molecular and cellular biology laboratory setting, exposing the students to a number of techniques that are commonly used in biochemistry, molecular and cellular biology research laboratories. These include enzyme kinetics, spectrophotometry, plasmid transformation, plasmid isolation, plasmid restriction digestion, protein expression and purification, small-interfering RNAs (siRNA) and gene expression, isolation of RNA, analysis of gene expression by polymerase chain reaction (PCR) and RT-qPCR, immunofluorescence and confocal microscopy. The course itself consists of several parts. The lectures, held prior to the beginning of a new experiment, serve to provide students with background on the experiment and/or techniques to be used, and may also explore alternative techniques. The students will then get to put into practice what they have learned from the lecture and reading the laboratory manual, as well as additional materials, if necessary. Finally, the laboratory report on each experiment allows the assessment of what the students have done, and also help train the students in proper experimental recording methods. All of these components will help the students to develop as a scientist.
Course Description from Program Guide	This course covers principles and practices of basic molecular and cellular biology techniques; Introduction to skills in a Molecular and Cellular Biology lab; Plasmids; transformation, isolation and restriction digestion; protein production, purification and functional assay; Small-interfering RNAs (siRNA) and gene expression; Isolation of RNA and Analysis of gene expression by Polymerase Chain Reaction (PCR) and RT-qPCR; Immunofluorescence; following cellular movement of a shuttling RNA-binding protein before and after stress.
Goals and Objectives	The course aim to provide first hands-on experience in a molecular and cellular biology laboratory setting, exposing the students to a number of techniques that are commonly used in biochemistry, molecular and cellular biology research laboratories. This will give MS students basic skills to start their research directed study courses, internships and thesis work.
Required Knowledge	The students are expected to have basic foundation in biochemistry, molecular biology and cell biology.

Reference Texts	Laboratory Manual for Methods in Molecular and Cellular Biology
Method of evaluation	85.00% - Homework /Assignments 15.00% - Active participation
Nature of the assignments	The assignments will consist of lab written reports, lab oral presentation reports, experimental flow sheets, short essay and lab quizzes
Course Policies	Attendance of and active participation in lectures and lab is mandatory. Any planned absence needs to be discussed with the course instructor.
Additional Information	

Tentative Course Schedule

(Time, topic/emphasis & resources)

Week	Lectures	Topic
1	Mon 08/27/2018 Thu 08/30/2018	Introduction to Molecular and Cellular Biology Experiments
2	Mon 09/03/2018 Thu 09/06/2018	Enzyme Kinetics/Spectrophotometry
3	Mon 09/10/2018 Thu 09/13/2018	Plasmids: Transformation, Isolation and Restriction Digestion
4	Mon 09/17/2018 Thu 09/20/2018	Protein Purification and Translation
5	Mon 09/24/2018 Thu 09/27/2018	Protein Purification and Translation
6	Mon 10/01/2018 Thu 10/04/2018	Evaluation
7	Mon 10/08/2018 Thu 10/11/2018	Small-interfering RNAs (siRNA) and gene expression
8	Mon 10/15/2018 Thu 10/18/2018	Evaluation
9	Mon 10/22/2018 Thu 10/25/2018	Isolation of RNA and Analysis of gene expression by Polymerase Chain Reaction (PCR) and RT-qPCR
10	Mon 10/29/2018 Thu 11/01/2018	Isolation of RNA and Analysis of gene expression by Polymerase Chain Reaction (PCR) and RT-qPCR
11	Mon 11/05/2018 Thu 11/08/2018	Evaluation
12	Mon 11/12/2018 Thu 11/15/2018	Immunofluorescence: following the cellular movement of a shuttling RNA-binding protein before and after stress
13	Mon 11/19/2018 Thu 11/22/2018	Immunofluorescence: following the cellular movement of a shuttling RNA-binding protein before and after stress
14	Mon 11/26/2018 Thu 11/29/2018	Evaluation
15	Mon 12/03/2018 Thu 12/06/2018	Evaluation
16	Mon 12/10/2018	Final Exam
17		
18		

Note

The instructor reserves the right to make changes to this syllabus as necessary.

Course Syllabus: Molecular & Cellular Biology - B 241

Division	Biological and Environmental Sciences & Engineering Division
Course Number	B 241
Course Title	Molecular & Cellular Biology
Academic Semester	Fall
Academic Year	2018/2019
Semester Start Date	08/26/2018
Semester End Date	12/11/2018
Class Schedule (Days & Time)	01:00 PM - 02:30 PM Sun Wed

Instructor(s)				
Name	Email	Phone	Office Location	Office Hours
Samir Hamdan	samir.hamdan@kaust.edu.sa	+966128082384		Tuesday: 10:00-11:00 am Building 2 Room 4221
Samah Zeinab Gadhoun	Samah.Gadhoun@kaust.edu.sa	+966128082930		

Teaching Assistant(s)	
Name	Email
Dr. Samah Zeineb Gadhoun; Building 9, third floor and room 3354	samah.gadhoun@kaust.edu.sa

Course Information	
Comprehensive Course Description	The course provides first hands-on experience in a molecular and cellular biology laboratory setting, exposing the students to a number of techniques that are commonly used in biochemistry, molecular and cellular biology research laboratories. These include enzyme kinetics, spectrophotometry, plasmid transformation, plasmid isolation, plasmid restriction digestion, protein expression and purification, small-interfering RNAs (siRNA) and gene expression, isolation of RNA, analysis of gene expression by polymerase chain reaction (PCR) and RT-qPCR, immunofluorescence and confocal microscopy. The course itself consists of several parts. The lectures, held prior to the beginning of a new experiment, serve to provide students with background on the experiment and/or techniques to be used, and may also explore alternative techniques. The students will then get to put into practice what they have learned from the lecture and reading the laboratory manual, as well as additional materials, if necessary. Finally, the laboratory report on each experiment allows the assessment of what the students have done, and also help train the students in proper experimental recording methods. All of these components will help the students to develop as a scientist.
Course Description from Program Guide	This course covers principles and practices of basic molecular and cellular biology techniques; Introduction to skills in a Molecular and Cellular Biology lab; Plasmids; transformation, isolation and restriction digestion; protein production, purification and functional assay; Small-interfering RNAs (siRNA) and gene expression; Isolation of RNA and Analysis of gene expression by Polymerase Chain Reaction (PCR) and RT-qPCR; Immunofluorescence; following cellular movement of a shuttling RNA-binding protein before and after stress.
Goals and Objectives	The course aim to provide first hands-on experience in a molecular and cellular biology laboratory setting, exposing the students to a number of techniques that are commonly used in biochemistry, molecular and cellular biology research laboratories. This will give MS students basic skills to start their research directed study courses, internships and thesis work.
Required Knowledge	The students are expected to have basic foundation in biochemistry, molecular biology and cell biology.
Reference Texts	Laboratory Manual for Methods in Molecular and Cellular Biology
Method of evaluation	60.00% - Homework /Assignments 40.00% - Oral presentation

Nature of the assignments	The assignments will consist of lab written reports, lab oral presentation reports, experimental flow sheets, short essay and lab quizzes.
Course Policies	Attendance of and active participation in lectures and lab is mandatory. Any planned absence needs to be discussed with the course instructor.
Additional Information	

Tentative Course Schedule

(Time, topic/emphasis & resources)

Week	Lectures	Topic
1	Sun 08/26/2018 Wed 08/29/2018	Introduction to Molecular and Cellular Biology Experiments
2	Sun 09/02/2018 Wed 09/05/2018	Enzyme Kinetics/Spectrophotometry
3	Sun 09/09/2018 Wed 09/12/2018	Plasmids: Transformation, Isolation and Restriction Digestion
4	Sun 09/16/2018 Wed 09/19/2018	Protein Purification and Translation
5	Sun 09/23/2018 Wed 09/26/2018	Evaluation/Tutorial
6	Sun 09/30/2018 Wed 10/03/2018	Evaluation/Tutorial
7	Sun 10/07/2018 Wed 10/10/2018	Small-interfering RNAs (siRNA) and gene expression
8	Sun 10/14/2018 Wed 10/17/2018	Evaluation/Tutorial
9	Sun 10/21/2018 Wed 10/24/2018	Isolation of RNA and Analysis of gene expression by Polymerase Chain Reaction (PCR) and RT-qPCR
10	Sun 10/28/2018 Wed 10/31/2018	Evaluation/Tutorial
11	Sun 11/04/2018 Wed 11/07/2018	Evaluation/Tutorial
12	Sun 11/11/2018 Wed 11/14/2018	Immunofluorescence: following the cellular movement of a shuttling RNA-binding protein before and after stress
13	Sun 11/18/2018 Wed 11/21/2018	Evaluation/Tutorial
14	Sun 11/25/2018 Wed 11/28/2018	Evaluation/Tutorial
15	Sun 12/02/2018 Wed 12/05/2018	Evaluation/Tutorial
16	Sun 12/09/2018	Final Exam
17		
18		

Note

The instructor reserves the right to make changes to this syllabus as necessary.